



Review

Adequate dietary vitamin D and calcium are both required to reduce bone turnover and increased bone mineral volume



Alice M.C. Lee^{a,b}, Rebecca K. Sawyer^{a,b}, Alison J. Moore^{a,b}, Howard A. Morris^{a,b}, Peter D. O'Loughlin^{a,b}, Paul H. Anderson^{a,b,*}

^a Centre for Musculoskeletal Health Research, School of Pharmacy and Medical Sciences, University of South Australia, Adelaide, SA, Australia

^b Chemical Pathology, SA Pathology, Adelaide, SA, Australia

ARTICLE INFO

Article history:

Received 1 August 2013

Received in revised form

11 November 2013

Accepted 13 November 2013

Available online 2 December 2013

Keywords:

Vitamin D

Calcium

Bone turnover

ABSTRACT

Clinical studies indicate that the combination of vitamin D and dietary calcium supplementation is more effective for reducing fracture risk than either supplement alone. Our previous dietary studies demonstrated that an adequate serum 25-hydroxyvitamin D₃ (25D) of 80 nmol/L or more reduces bone RANKL expression, osteoclastogenesis and maintains the optimal levels of trabecular bone volume (BV/TV%) in young rats. The important clinical question of the interaction between vitamin D status, dietary calcium intake and age remains unclear. Hence, 9 month-old female Sprague-Dawley rats ($n = 5-6/\text{group}$) were pair-fed a semi-synthetic diet containing varying levels of vitamin D (0, 2, 12 or 20 IU/day) and dietary calcium (0.1% or 1%) for 6 months. At 15 months of age, animals were killed, for biochemical and skeletal analyses. While changes to serum 25D were determined by both dietary vitamin D and calcium levels, changes to serum 1,25-dihydroxyvitamin D₃ (1,25D) were consistently raised in animals fed 0.1% Ca regardless of dietary vitamin D or vitamin D status. Importantly, serum cross-laps levels were significantly increased in animals fed 0.1% Ca only when combined with 0 or 2 IUD/day of vitamin D, suggesting a contribution of both dietary calcium and vitamin D in determining bone resorption activity. Serum 25(OH)D₃ levels were positively correlated with both femoral mid-diaphyseal cortical bone volume ($R^2 = 0.24$, $P < 0.01$) and metaphyseal BV/TV% ($R^2 = 0.23$, $P < 0.01$, data not shown). In multiple linear regressions, serum 1,25(OH)₂D₃ levels were a negative determinant of CBV ($R^2 = 0.24$, $P < 0.01$) and were not a determinant of metaphyseal BV/TV% levels. These data support clinical data that reduced bone resorption and increased bone volume can only be achieved with adequate 25D levels in combination with high dietary calcium and low serum 1,25D levels.

This article is part of a Special Issue entitled '16th Vitamin D Workshop'.

Crown Copyright © 2013 Published by Elsevier Ltd. All rights reserved.

Contents

1. Introduction	160
2. Materials and methods	160
2.1. Animals	160
2.2. Biochemical analyses	160
2.3. Micro-computed topographical analyses	160
2.4. Data expression and statistical analyses	160
3. Results	160
3.1. Serum biochemistry	160
3.2. Bone mineral analyses	160
3.3. Relationships between biochemistry and bone structure	161
4. Discussion	161
References	161

* Corresponding author at: Centre for Musculoskeletal Health Research, School of Pharmacy and Medical Sciences, University of South Australia, Adelaide, SA 5000, Australia. Tel.: +61 8 82223244; fax: +61 8 82223518.

E-mail addresses: paul.anderson@unisa.edu.au, paul.anderson@health.sa.gov.au (P.H. Anderson).

1. Introduction

Osteoporosis is a preventable bone disease which can occur as a consequence of prolonged depletion of circulating levels of 25 hydroxyvitamin D (25D). The level of 25D required in humans to prevent osteoporosis has been reported to be approximately 75 nmol/L which is necessary to maintain normal bone mineral density and reduce the incidence of hip fracture in the elderly as determined by meta-analyses of clinical trials when vitamin D supplementation is 800 IU per day or greater in combination with calcium supplementation [1,2]. We have previously reported that vitamin D depletion in rats causes osteopenia in at least three skeletal sites including the distal and proximal femoral metaphyses and vertebrae. Furthermore, serum 25D levels ranging from 20 to 115 nmol/L were a positive and independent determinant of femoral trabecular bone volume [3]. However, the level of vitamin D required to prevent bone loss is likely to be dependent on factors such as age and nutrition and in particular the level of dietary calcium intake [4]. The co-administration of vitamin D and calcium reduces the incidence of hip fracture in elderly [5]. Numerous clinical trials demonstrate that vitamin D supplementation sufficient to raise serum 25D levels has greater efficacy in reducing fracture rates if combined with calcium supplementation [5,6]. It is clear that a diet containing high levels of calcium protects against bone loss by reducing both parathyroid hormone secretion and 1,25-dihydroxyvitamin D₃ (1,25D) production, which in turn reduces osteoclastic activity and anti-mineralisation [7,8]. We and others have reported that circulating levels of serum 25D are determined by both dietary vitamin D and calcium in both an animal model [9] and in postmenopausal women [10] suggesting that high dietary calcium may also play a role maintaining serum 25D levels.

In order to further understand the relationship between serum 25D levels and bone health, we have utilised our previously developed animal model [9] to address the question as to whether dietary calcium restriction influences the relationship between vitamin D status and bone mineral volume.

2. Materials and methods

2.1. Animals

Nine-month old female Sprague-Dawley rats ($n=5-6$ /group) were allocated to a diet containing varying levels of vitamin D₃ (0, 2, 12 or 20 IU/day) in combination with either 0.1% or 1% calcium, based on the recommended semi-synthetic diet for rodents (AIN-93-VX) [11]. All animals were maintained on this diet for 6 months, at which time animals were killed. All animal procedures were approved by the Institutional Animal Ethics Committee.

2.2. Biochemical analyses

Food was withdrawn from animals 16 h before blood samples were collected via the tail vein at time of death. Serum 1,25D and 25D levels were measured by RIA (Immunodiagnostic Systems Ltd.,

Bolden, UK). Levels of serum C-terminal telopeptide α 1 chain of rat type I collagen (Cross-laps) were measured by EIA (Immunodiagnostic Systems Ltd., Bolden, UK). Serum calcium and inorganic phosphate, were measured using a chemistry analyser (Trace Scientific reagents, Vic, Australia; Hitachi 911 automated analyser, Roche, IN, USA).

2.3. Micro-computed topographical analyses

The proximal femur and tibia bone micro-architecture was analysed using a high-resolution micro-CT system (Skyscan 1072, Belgium) to obtain multiple X-ray transmission images. The resolution of scanning was 6.5 μ m/pixel. Cross sectional images of the object were then reconstructed by a modified Feldkamp cone-beam algorithm, creating a complete 3D representation of internal microstructure and density. Adaptive thresholding technique was applied to reconstructed dataset prior to calculations of metaphyseal BV/TV%, Tb.N and Tb.Th. For cortical bone, an 8 mm mid-region of the femur was used to determine cortical bone volume.

2.4. Data expression and statistical analyses

The effects of diet on histological measurements and biochemical markers were statistically analysed by multivariate analysis of variance and Tukey's post hoc test analysis. Multiple linear regression analyses were used to identify determinants of bone mineral volume. A P value of 0.05 or less was considered to be statistically significant.

3. Results

3.1. Serum biochemistry

Serum 25D levels were lowest in the rats fed no vitamin D and 0.1% calcium diet (Table 1). Serum 25D levels were significantly greater in animals fed 1% calcium in the 0, 2 and 20 IU vitamin D₃/day fed animals when compared to animals fed 0.1% dietary calcium ($P<0.05$). The animals fed 20 IU vitamin D₃/day and 1% calcium recorded the highest mean serum 25D ($P<0.05$). Despite the relative low serum 25D levels in animal fed 0 IU vitamin D₃/day and 0.1% calcium, serum 1,25D levels were significantly increased and comparable to levels in vitamin D-replete animals fed 0.1% calcium. Mean serum calcium levels were significantly elevated in the animals fed 20 IU vitamin D₃/day and 1% calcium. Serum cross-laps were highest in rats fed 0.1% Ca and receiving 0 or 2 IU vitamin D₃/day.

3.2. Bone mineral analyses

While all animals fed 1% calcium demonstrated higher metaphyseal BV/TV% of the distal femur, the highest BV/TV% was in rats fed either 12 or 20 IU vitamin D₃/day. The higher metaphyseal BV/TV% in 1% calcium fed rats was due to increased Tb.Th, except for the rats fed 20 IU vitamin D₃/day, where increased BV/TV% was due to both

Table 1
Serum levels of 25 hydroxyvitamin D, 1,25-dihydroxyvitamin D, calcium and cross-laps in rats fed varying levels of vitamin D and either 0.1% or 1% dietary calcium.

Vitamin D (IU/day)	0		2		12		20	
	0.1	1	0.1	1	0.1	1	0.1	1
Serum 25D (nmol/L) (sem)	21.7 (2.9)	84.5 (6.5) ^a	69.5 (9.4)	86.2 (7.8)	87.8 (10.7)	119.4 (11.6) ^a	90.9 (11.8)	161.3 (38.8) ^a
Serum 1,25D (pmol/L) (sem)	246.6 (30.2)	19.5 (6.5) ^a	226 (32.4)	48.0 (15.3) ^a	216.6 (31.8)	9.0 (4.5) ^a	248.6 (68.1)	13.8 (3.5) ^a
Serum Ca (mmol/L) (sem)	2.8 (0.1)	2.6 (0.1)	2.7 (0.2)	2.5 (0.1)	2.9 (0.2)	2.8 (0.2)	2.6 (0.2)	3.2 (0.2) ^a
Serum X-laps (nmol/L) (sem)	20.8 (2.4)	13.0 (1.5) ^a	21.7 (9.0)	6.0 (4.0) ^a	10.5 (2.1)	6.7 (2.2)	7.8 (2.0)	9.6 (5.1)

Values are mean (sem), $n=5-6$. 25D: 25-hydroxyvitamin D₃; 1,25D: 1,25 dihydroxyvitamin D₃; Ca: calcium; X-laps, c-telopeptide fragments of collagen-type I.

^a $P<0.05$ vs. 0.1% Ca groups.

Table 2
Static bone histomorphometry of the distal femoral metaphysis.

Vitamin D (IU/kg)	0		2		12		20	
	0.1	1	0.1	1	0.1	1	0.1	1
BV/TV (%) (sem)	22.6 (1.8)	27.0 (1.8) ^a	26.3 (1.6)	26.9 (0.5) ^a	25.1 (0.9)	29.3 (1.0) ^a	25.8 (1.4)	29.3 (1.7) ^a
Tb.N (mm) (sem)	2.65 (0.1)	2.74 (0.12)	2.75 (0.12)	2.82 (0.05)	2.71 (0.13)	2.93 (0.07)	2.74 (0.11)	3.12 (0.11) ^a
Tb.Th (μm) (sem)	79.2 (3.1)	88.3 (1.5) ^a	82.7 (2.9)	87.0 (1.7) ^a	83.3 (2.1)	91.4 (2.3) ^a	86.6 (0.8) ^b	91.7 (2.5) ^a
Cort BV (mm ³) (sem)	27.2 (0.8)	28.6 (0.6)	27.3 (1.1)	28.8 (0.4)	27.3 (1.4)	30.8 (1.2) ^a	27.1 (0.9)	31.7 (1.2) ^a

Values are mean (sem), *n* = 6. BV/TV: bone volume/total volume; Tb.N: trabecular number; Tb.Th: trabecular thickness.

^a *P* < 0.05 vs. 0.1% Ca groups.

^b *P* < 0.05 vs. 0 IU Vit D/0.1% Ca.

Table 3
Dynamic bone histomorphometry of the distal femoral metaphysis.

Vitamin D (IU/kg)	0		2		12		20	
	0.1	1	0.1	1	0.1	1	0.1	1
MS/BS (sem)	29.5 (0.8)	32.4 (3.0)	32.4 (2.9)	33.7 (4.1)	33.8 (2.2)	25.7 (2.5) ^a	27.7 (1.8)	17.9 (1.7) ^a
MAR, (μm/day) (sem)	1.22 (0.05)	1.19 (0.06)	1.25 (0.04)	1.23 (0.01)	1.37 (0.04)	1.37 (0.04)	1.22 (0.02)	1.22 (0.06)
BFR (μm ³ /μm ² μday) (sem)	36.1 (1.6)	38.5 (3.8)	41.0 (4.7)	40.0 (1.3)	45.9 (2.3)	35.0 (3.1) ^a	33.9 (3.4)	21.8 (2.3) ^a
Oc/B.Pm, no. 10 ⁻³ × (mm) (sem)	3.1 (0.7)	2.0 (0.4)	1.8 (0.2) ^b	1.4 (0.2)	2.1 (0.4)	1.4 (0.3) ^a	2.0 (0.2)	1.1 (0.1) ^a

Values are mean (sem), *n* = 6. MS/BS: mineralising surface/bone surface; MAR: mineral apposition rate; BFR: bone formation rate; Oc/B.Pm: osteoclast number/bone perimeter.

^a *P* < 0.05 vs. 0.1% Ca groups.

^b *P* < 0.05 vs. 0 IU Vit D/0.1% Ca.

Table 4
Multiple linear regression equations for serum 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D as determinants of femoral cortical bone volume of the mid-diaphyseal region.

Independent variable	Equation	R ²	<i>P</i> value
Serum 25D + Serum 1,25D	CBV = +0.016 × 25D − 0.008 × 1,25D + 28.4	0.24 0.28	0.005 0.02
Multiple R ²		0.33	

CBV: cortical bone volume; 25D: 25 hydroxyvitamin D3; 1,25D: 1,25 dihydroxyvitamin D3.

Th.Th and Tb.N. Similar to BV/TV% measures, cortical bone volume was greatest in rats fed 12 and 20 IU vitamin D₃/day plus 1% Ca.

3.3. Relationships between biochemistry and bone structure

Serum 25D levels were positively correlated with both femoral mid-diaphyseal cortical bone volume (*R*² = 0.24, *P* < 0.01) (Table 4) and metaphyseal BV/TV% (*R*² = 0.23, *P* < 0.01, data not shown). In multiple linear regressions, serum 1,25D levels were a negative determinant of CBV (*R*² = 0.24, *P* < 0.01) (Table 4) and were not determinant of metaphyseal BV/TV% levels (Tables 2 and 3).

4. Discussion

The association between circulating 25D levels and bone mineral volume at two skeletal sites confirms our previous report that serum 25D levels are an independent predictor of bone mineral volume [3]. Previously, we reported that serum 25D levels were determined by both dietary vitamin D and calcium levels [9], suggesting that the benefits of high dietary calcium in improving bone mineral volume may be, in part, by preserving serum 25D levels. All animals fed the low 0.1% Ca diet demonstrated markedly increased serum 1,25D levels, which were a negative determinant of cortical bone volume, consistent with the reported roles for high circulating 1,25D levels in stimulating bone resorption and inhibition of mineralisation [12,13]. However, raised levels of both osteoclast numbers within the metaphysis and serum cross-laps occurred with higher serum 1,25D levels only when serum 25D levels were reduced, suggesting that 25D may also play a role in directly regulating the bone resorption response, as we have previously observed in *in vitro*

studies [14,15]. Conversely, lower levels of mineralising surface and bone formation rate only occurred in those animals with levels of serum 25D above 90 nmol/L levels and with low 1,25D levels. This suggests that increased bone volume, associated with adequate vitamin D and dietary calcium, is due to reduced bone turnover which favours maintenance of bone mineral levels.

We have previously reported that 25D directly promotes bone mineralisation, through its conversion to 1,25D within bone cells [16,17]. It is plausible that vitamin D-depletion may directly impair local synthesis of 1,25D in bone cells and thus bone mineralisation. How local synthesis of 1,25D in bone cells contributes to changes in bone volume in the context of changes to circulating 1,25D remains to be fully elucidated. However, partial depletion of 25D, due to seasonal changes have been previously shown in human studies not to be associated with changes in intestinal calcium absorption [18], and yet both osteoid volume and mineralisation lag time were increased, highlighting that the association between serum 25D levels and bone mineral may be due to altered local synthesis of 1,25D [18].

Given the several reports of VDR-mediated activities in osteoblasts, osteocytes and osteoclast, as well as chondroblasts, which appear to regulate both anabolic and catabolic processes depending on the physiological context, data from this study support the notion that both 25D and 1,25D facilitate calcium flow in and out of the skeleton presumably to primarily preserve normocalcemia with both positive and negative effects on the skeleton.

References

- [1] H.A. Bischoff-Ferrari, et al., Fracture prevention with vitamin D supplementation: a meta-analysis of randomized controlled trials, *JAMA* 293 (18) (2005) 2257–2264.
- [2] H.A. Bischoff-Ferrari, et al., Estimation of optimal serum concentrations of 25-hydroxyvitamin D for multiple health outcomes, *Am. J. Clin. Nutr.* 84 (1) (2006) 18–28.
- [3] P.H. Anderson, et al., Vitamin D depletion induces RANKL-mediated osteoclastogenesis and bone loss in a rodent model, *J. Bone Miner. Res.* 23 (11) (2008) 1789–1797.
- [4] R.L. Bailey, et al., Estimation of total usual calcium and vitamin D intakes in the United States, *J. Nutr.* 140 (4) (2010) 817–822.
- [5] M.C. Chapuy, et al., Vitamin D3 and calcium to prevent hip fractures in the elderly women, *N. Engl. J. Med.* 327 (23) (1992) 1637–1642.
- [6] M.C. Chapuy, et al., Combined calcium and vitamin D3 supplementation in elderly women: confirmation of reversal of secondary hyperparathyroidism

- and hip fracture risk: the Decalys II study, *Osteoporosis Int.* 13 (3) (2002) 257–264.
- [7] M. Yamaguchi, M.N. Weitzmann, High dose 1,25(OH)₂D₃ inhibits osteoblast mineralization in vitro, *Int. J. Mol. Med.* 29 (5) (2012) 934–938.
- [8] J.S. Adams, G. Lee, Gains in bone mineral density with resolution of vitamin D intoxication, *Ann. Int. Med.* 127 (3) (1997) 203–206.
- [9] P.H. Anderson, et al., The effect of dietary calcium on 1,25(OH)₂D₃ synthesis and sparing of serum 25(OH)D₃ levels, *J. Steroid Biochem. Mol. Biol.* 121 (1/2) (2010) 288–292.
- [10] S.D. Thomas, A.G. Need, B.E. Nordin, Suppression of C-terminal telopeptide in hypovitaminosis D requires calcium as well as vitamin D, *Calcif. Tissue Int.* 86 (5) (2010) 367–374.
- [11] P.G. Reeves, F.H. Nielsen, G.C. Fahey Jr., AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet, *J. Nutr.* 123 (11) (1993) 1939–1951.
- [12] M.H. Lafage-Proust, et al., High bone turnover persisting after vitamin D repletion: beware of calcium deficiency, *Osteoporos Int.* 24 (8) (2013) 2359–2363.
- [13] L. Lieben, et al., Normocalcemia is maintained in mice under conditions of calcium malabsorption by vitamin D-induced inhibition of bone mineralization, *J. Clin. Invest.* 122 (5) (2012) 1803–1815.
- [14] M. Kogawa, et al., The metabolism of 25(OH)-vitamin D₃ by osteoclasts and their precursors regulates the differentiation of osteoclasts, *J. Steroid Biochem. Mol. Biol.* 121 (2010) 277–280.
- [15] M. Kogawa, et al., Osteoclastic metabolism of 25(OH)-vitamin D₃: a potential mechanism for optimization of bone resorption, *Endocrinology* 151 (10) (2010) 4613–4625.
- [16] E.M. Gardiner, et al., Increased formation and decreased resorption of bone in mice with elevated vitamin D receptor in mature cells of the osteoblastic lineage, *FASEB J.* 14 (13) (2000) 1908–1916.
- [17] G.J. Atkins, et al., Metabolism of vitamin D(3) in human osteoblasts: evidence for autocrine and paracrine activities of 1alpha,25-dihydroxyvitamin D(3), *Bone* 40 (6) (2007) 1517–1528.
- [18] A.G. Need, et al., Seasonal change in osteoid thickness and mineralization lag time in ambulant patients, *J. Bone Miner. Res.* 22 (5) (2007) 757–761.