

# Probiotics: Mechanisms of Action and Clinical Applications

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## Abstract

Probiotics are living microorganisms which when taken in adequate amount provides benefit to the host. While this beneficial effect was originally thought to stem from improvements in the intestinal microbial balance, there is now substantial evidence that probiotics can also provide benefits by modulating immune functions. Extrapolation of immunomodulatory effects found in the laboratory and in animal studies with outcomes in human trials presents a difficult challenge. Not all probiotics are created equal and the benefits are strain and dose specific. With newer strain-specific clinical trials and meta-analysis of the clinical trials, the beneficial role of probiotics in certain diseases has been evolving. Some uncertainty still exists with probiotics in other diseases with regard to the therapeutic role, strain-specificity, dosage and duration. Identification of clinical characteristics of effective probiotic strains, their mechanisms of action and testing of probiotic-based treatment may provide the true beneficial effect of probiotics in various disorders.

**Keywords:** Probiotics; Bacteriocins; Microcins; Antibiotic-associated diarrhea; *Clostridium difficile* infection; Acute pancreatitis; Necrotizing enterocolitis; Ventilator-associated pneumonia, Multi-organ dysfunction syndrome

## Introduction

Recent research has revealed a potential therapeutic role for the manipulation of the microbiota in the maintenance of human health and treatment of various mucosal disorders. Probiotic microorganisms can shape the immune system both at the local and systemic level which will allow future probiotics as treatments for many diseases. The benefits include either a shortened duration of infections or decreased susceptibility to pathogens [1].

The word 'probiotic', derived from the Greek language, means 'for life' was first used by Kollath [2]. Lilly and Stillwell [3] defined probiotics as substances produced by microorganisms which promoted the growth of other microorganisms. According to the currently adopted definition by FAO/WHO [4], probiotics are: "Live microorganisms which when administered in adequate amounts confer a health benefit on the host". Prebiotics are indigestible food ingredients that selectively promote the growth or activity of beneficial bacteria, thereby benefiting the host [5]. Synbiotics are combinations of probiotics and prebiotics designed to improve the survival of ingested microorganisms and their colonization of the intestinal tract [5]. Commonly used bacterial probiotics include *Lactobacillus* species, *Bifidobacterium* species, *Escherichia coli*, *Streptococcus* species, *Lactococcus lactis* and some *Enterococcus* species. Currently, the only probiotic yeast used is the nonpathogenic *Saccharomyces boulardii*.

## Mechanisms of Probiotic Function

Probiotics do not always colonize the intestinal tract to exert their effects. For example, some probiotics like *Bifidobacterium longum* become part of the human intestinal microflora, whereas others like *Lactobacillus casei* indirectly exert their effects in a transient manner as they pass through by remodeling or influencing the existing microbial community [6]. The following are the major mechanisms of action of probiotics on the host (Table 1).

## Barrier function

Probiotics are capable of influencing many of the components of epithelial barrier function either by decreasing apoptosis of intestinal cells or increased mucin production. *Lactobacillus rhamnosus* GG was able to prevent cytokine-induced apoptosis in intestinal epithelial cell models by inhibiting tumor necrosis factor (TNF) [7]. *Lactobacillus* species have been shown to increase mucin expression *in vitro* in human intestinal epithelial cells, thus blocking pathogenic *E. coli* invasion and adherence [8,9]. *Lactobacillus rhamnosus* GG has shown to prevent inflammation and programmed cell death of the lining intestinal epithelial cells [10] and shown to exert mitogenic effects and enhancing mucosal regeneration [11].

## Production of antimicrobial substances

Probiotics either by inducing host cells to produce peptides or by directly releasing peptides interfere with pathogens, and prevent epithelial invasion. Defensins (hBD protein) and cathelicidins are the antimicrobial peptides expressed constitutively by the intestinal epithelial cells and display antimicrobial activity against a wide variety of bacteria, fungi and some viruses [12]. Certain probiotic strains like *E. coli* strain DSM 17252 G2 (one of the three Symbioflor 2 genotype strains) and several *Lactobacilli* species have shown to express certain defensins [13]. Healthy volunteers who received probiotics had increased fecal hBD protein and remained elevated for 9 weeks after completion of 3 weeks of probiotic treatment [13,14]. Probiotics have been shown to suppress pathogen growth through the release of a variety of antimicrobial factors like defensins, bacteriocins,

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Mode	Process	Mechanism	Examples
Barrier Function	Decreased apoptosis of epithelial cells	Decreased TNF- $\alpha$ production	<i>Lactobacillus rhamnosus</i> GG
	Increasing mucin production	Increased expression of MUC 2	<i>Lactobacillus</i> sp
Host cell Antimicrobial Peptides	Defensins (hBD protein)	Increased up regulation of Defensin	<i>E coli strain</i> DSM 17252S2
	Cathelicidins	By butyrate production	
Probiotic Antimicrobial Factors	Lowering the luminal pH	By secretion of SCFA's	Most of the probiotics bacteria
	Bacteriocin production	By Gram positive probiotics	
	Microcin production	By Gram negative probiotics	
Epithelial Adherence	By competing with pathogens	Directly or indirectly by producing protein that block adherence	
Immune Modulation	Blocking pro Inflammatory molecules	By attenuating IL-8 secretion or blocking the degradation of the counter-regulatory factor I $\kappa$ B	<i>Salmonella typhimurium</i> VSL#3 probiotics
	Increasing mucosal immunity	Increasing IgA Production	<i>L. casei</i>
Interference with Quorum Sensing Signaling	Blocks the communication between pathogenic bacteria	By secreting molecules which blocks quorum sensing signaling	<i>L. acidophilus</i>

**Table 1:** Various mechanisms of probiotics action on human intestine cells.

Abbreviations: TNF- $\alpha$ : Tumor necrosis factor alpha; MUC 2: Mucin 2, hBD: Hemoglobin subunit delta; SCFA: Short chain fatty acids; IL-8: Interleukin 8; and I $\kappa$ B: Inhibitor of kappa B.

hydrogen peroxide, nitric oxide, and short chain fatty acids (SCFA), such as lactic and acetic acids, which reduce the pH of the lumen [15]. SCFA can disrupt the outer membranes of gram-negative pathogens causing inhibition of pathogen growth [16]. Bacteriocins can either permeabilize the inner membrane of gram-negative bacteria, leading to disruption and formation of pores [17]. Microcins (produced by gram negative bacteria), on the other hand, can target the inner membrane, enzymes that are involved in DNA or RNA structure and synthesis, or protein synthesis enzymes [18].

### Competition for adherence

Probiotic bacteria compete with invading pathogens for binding sites to epithelial cells and the overlying mucus layer in a strain-specific manner. Surface layer proteins purified from *L. helveticus* R0052 inhibited enterohemorrhagic *Escherichia coli* O157:H7 adherence and the subsequent rise in permeability, without altering the growth of the pathogen [19]. *S. bouardii* secretes a heat-labile factor which has shown to be responsible for the decreased bacterial adherence [20].

### Immune modulation

*L. casei* have been shown to augment total and pathogen-specific secretory IgA levels upon infection in mice by stimulating B cell class switching to IgA [21]. Specific antibodies against *L. casei* were not produced, indicating the non-responsiveness of the gut immune system to this beneficial bacterium. In infant rabbits pretreated with *L. casei*, morbidity of subsequent EHEC (Enterohemorrhagic *E. coli*) infection was reduced due to increased mucosal levels of anti-EHEC and anti-Shiga toxin IgA antibodies compared with controls [22]. *L. casei* down-regulated the transcription of a number of genes encoding pro-inflammatory effectors such as cytokines and chemokines and adherence molecules induced by invasive *S. flexneri*. This resulted in an anti-inflammatory effect that appeared mediated by the inhibition of the NF- $\kappa$ B pathway, particularly through stabilization of I- $\kappa$ B $\alpha$  [23].

### Interference with quorum sensing signaling

Bacteria communicate with each other as well as with their surrounding environment through chemical signalling molecules called auto-inducers. This phenomenon is called quorum sensing [24]. The use of this cell-to-cell signaling mechanism facilitates the regulation of important traits of enteric microbes that allow them to successfully colonize and/or start infection in their host [25]. Medellin-Pena et al. [26] demonstrated that *Lactobacillus acidophilus* secretes a molecule

that inhibits the quorum sensing signalling or directly interact with bacterial transcription of *E. coli* O157 gene, involved in colonization and thus, bacterial toxicity is opposed.

### Role of Probiotics in Various Diseases

Probiotic research is moving forward on two stages: laboratory studies and clinical trials to evaluate the safety and efficacy of probiotics in treatment and prevention of various medical conditions. In this article, we will review the evidence of probiotics in various diseases by reviewing the clinical trials and meta-analysis of the clinical trials (Table 2).

### Antibiotic-associated Diarrhea

The incidence of antibiotic-associated diarrhea (AAD) ranges between 5% and 30% [27]. The risk is greatest with aminopenicillin therapies (Ampicillin or Amoxicillin), aminopenicillin combined with clavulanic acid, cephalosporins, and clindamycin [27]. Probiotics given in conjunction with antibiotics have been extensively studied for the prevention of AAD in both adults and children. The major changes in the microbiota of the gut with antibiotics are decrease in total number and species diversity of *Bacteroides* and *Bifidobacteria* associated with decreased amyolytic activity with increase in facultative anaerobes such as *Fusobacteria*, *Clostridia*, and *Eubacteria* species [28]. Decreased short chain fatty acid production and increased proteolytic activity was also noted in elderly patients treated with antibiotics [29].

Several clinical trials have been conducted using *Saccharomyces bouardii* for the prevention of AAD [30-36]. All but one concluded that *S. bouardii* was an effective agent for prevention of AAD [34]. With increasing number of trials over the last several years on the role of probiotics in preventing AAD, new single-strain meta-analysis are now being published. A meta-analysis of several randomized controlled trials testing the efficacy of *S. bouardii* in preventing AAD in adults showed *S. bouardii* was significantly protective for AAD with an overall pooled relative risk of 0.47 (95% Confidence Interval=0.35, 0.63; p<0.001) [37]. The number needed to prevent one case of AAD was 10.2. A meta-analysis of five randomized controlled trials by Szajewska and Mrukowicz [38] involving 1076 subjects showed a significantly protective effect of *S. bouardii* was found (pooled RR=0.43, 95% CI: 0.23-0.78).

In addition, several randomized controlled trials have shown efficacy for *Lactobacillus rhamnosus* GG (LGG) in prevention of AAD

[39-42]. Two of these trials focused on children only [40,41]. All but one showed a benefit over placebo or no treatment [42].

*Saccharomyces boulardii* alone is available as a probiotic in the market and sold as Florastor capsules. It contains 5 billion colony forming units. *Lactobacillus rhamnosus* GG (LGG) alone is available in market as Culturelle capsules and it contains 10 billion CFU's in each capsule (Table 3).

### Probiotics and *Clostridium difficile* Infections

*Clostridium difficile* is a spore-forming, anaerobic, Gram-positive bacterium that causes gastrointestinal infection with diarrhea and colitis. There has been a marked increase in the incidence and severity of *Clostridium difficile* infection (CDI) during the past decade. The clinical outcomes of CDI range from asymptomatic carriage to mild diarrhea to fulminant, often fatal, pseudomembranous colitis. Recurrent CDI is one of the most challenging aspects of the disease. Approximately 25% of patients treated for CDI with metronidazole or vancomycin experience recurrent symptoms, typically within 4 weeks of completing antibiotic therapy. Owing to increasing incidence, rising death rates, and frequent recurrences, there is a substantial need for more effective approaches to CDI prevention and therapy.

Castagliuolo et al. [43] found a 54 kDa serine protease produced by *S. boulardii* which directly degrades *C. difficile* toxin A and B and also produces a protease capable of degrading the colonic receptor site for *C. difficile*. *S. boulardii* may cause an increase in anti-toxin secretory IgA levels in the intestine [44]. Probiotics have been studied in prevention, and treatment of *Clostridium difficile* infections (CDI) and recurrent CDI.

Several randomized-controlled trials used *Lactobacillus* spp, *Saccharomyces boulardii* or a combination with *C. difficile* toxin acquisition and/or CDI as a primary or secondary outcome [45-52]. The trials had a small number of cases and short follow-up, the longest being 7 weeks by McFarland et al. [45]. Hickson et al. [52] showed a

statistically significant decrease in CDI with use of a combination probiotic milkshake. No patients in the probiotic group acquired CDI, whereas 9 out of 53 (17%) in the placebo group developed CDI (P=0.001). None of the remaining trials demonstrated a statistically significant decrease in CDI or *C. difficile* toxin acquisition with the use of probiotic therapy [45-51]. The above trials lacked adequate statistical power to determine the efficacy of probiotics. Few studies of probiotics have been performed but none has shown a consistent evidence of efficacy in prevention or treatment of CDI.

In another randomized, controlled trial, patients with recurrent CDI were prescribed either one of two doses of vancomycin (2 g/d or 500 mg/d) or metronidazole (1 g/d) then randomized to either *S. boulardii* or placebo (1 g/d for 4 weeks). Patients treated with the high dose vancomycin and the probiotic had significantly decreased recurrence rates (16.7%) compared to vancomycin and placebo (50%) [53]. The probiotic given with the low dose vancomycin or metronidazole was not significantly protective of CDI. *S. boulardii* was shown to be effective in recurrent CDI.

*Saccharomyces boulardii* alone is available in the market as Florastor capsules. *Lactobacillus* spp alone are available as Lactinex, Fem-Dophilus, and Culturelle capsules. *Lactobacillus* spp are available in combination with *Bifidobacterium* spp as Align capsules, Attune nutrition bars, Adult Formula CP-1 capsules, and OWP probiotics capsules. There has been increase in the practice of using probiotics along with vancomycin or metronidazole for recurrent CDI.

### Probiotics and *Helicobacter pylori* Infections

*Helicobacter pylori*, a small curved to spiral rod shaped bacterium, is strongly associated with duodenal peptic ulceration and it is the main etiologic agent of chronic gastritis and gastric cancer and other gastric malignancies. Today the therapy to eradicate this bacterium is based on a combination of antibiotics and proton pump inhibitors. Probiotics seem to have a direct antimicrobial effect, as shown through *in vitro*

Disease	Probiotic Strain	Comments
Prevention of antibiotic associated diarrhea (AAD)	<i>S. boulardii</i> [37,38]	Number needed to treat (NNT) is 10.2 to prevent one case of AAD.
	<i>Lactobacillus rhamnosus</i> GG (LGG) [39-41]	Effective on adults and children in RCT's.
Prevention of <i>Clostridium difficile</i> Infection (CDI)	<i>S. boulardii</i> , LGG, or both [52]	Study results are not statistically significant.
Prevention of recurrence after first CDI	<i>S. boulardii</i> [53]	Reduction of recurrence of CDI by half.
<i>Helicobacter pylori</i> eradication	<i>Lactobacillus rhamnosus</i> GG (LGG), <i>S. boulardii</i> , <i>L acidophilus</i> [55]	Moderate evidence for improving eradication but good evidence of reduction in side effects leading to improved compliance.
Ulcerative Colitis	<i>E coli</i> Nissle 1917 [68]	Promising role in maintenance of remission.
	VSL#3 [70]	Role in induction and maintenance of remission of UC.
Crohn's Disease	<i>Lactobacillus rhamnosus</i> GG (LGG) [71,72], <i>Lactobacillus johnsonii</i> LA1[73]	No role in induction or prolonging of remission of CD.
Irritable Bowel Syndrome	<i>Bifidobacterium infantis</i> 35624 [80], VSL#3 [81]	Significant improvements in IBS symptoms.
Acute Pancreatitis	<i>Lactobacillus plantarum</i> 299 [88]	But PROPATRIA trial showed an increased incidence of infection, MODS and bowel ischemia.
Necrotizing Enterocolitis (NEC)	<i>Bifidobacterium</i> spp and <i>Lactobacillus acidophilus</i> [148]	Prophylactic probiotics reduced NEC and mortality. Increased infection is noted among VLBW (<750 g).
Multi-Organ Dysfunction Syndrome (MODS)	VSL#3 [118]	Increased systemic IgA and IgG concentrations are noted but MODS scores were not significantly affected by probiotic treatment.
Allergy and Immune Response	<i>Lactobacillus rhamnosus</i> GG (LGG) [125-127]	Alone when given to mothers during pregnancy did decrease the risk of atopic dermatitis but similar results were not seen when given with other probiotic strains.
Ventilator Associated Pneumonia (VAP)	<i>Lactobacillus rhamnosus</i> GG (LGG) [142]	Number needed to treat (NNT) is 5 to prevent one case of VAP.

Table 2: List of different probiotic strains studied in treatment and/or prevention of various diseases.

studies, through competition with *H. pylori*, inhibition of adherence and production of metabolites and antimicrobial molecules.

In a randomized, double blind, placebo-controlled trial, 60 participants were treated with triple antibiotic therapy on days 1-7 and *Lactobacillus GG* on days 1-14 [54]. Probiotics significantly improved symptoms, including nausea, taste disturbance, and diarrhea; however, epigastric pain did not significantly improve during eradication treatment. Eradication rates did not differ significantly between the groups (83.3% vs 80%).

In another randomized, double blind, placebo-controlled trial, 85 *H. pylori* positive asymptomatic patients were randomized to receive *Lactobacillus GG* (group I), *Saccharomyces boulardii* (group II), *Lactobacillus acidophilus* and *Bifidobacterium lactis* (group III), or placebo (group IV) on days 1-14, with *H. pylori* treatment on days 1-7 [55]. Probiotics significantly improved symptoms, including taste disturbance and diarrhea; however, nausea and epigastric pain did not significantly improve during *H. pylori* treatment. All of the differences were noted between the probiotics and placebo. None of the probiotics was superior to another. Eradication rates were not significantly different among the 4 groups receiving probiotics.

Myllyluoma et al. [56] did a randomized double-blind, placebo-control trial on 47 patients using a milk based fruit drink containing *Lactobacillus GG*, *Propionibacterium*, *Bifidobacterium*, or placebo on days 1-28 and triple antibiotic therapy days 1-7. Probiotics did not significantly improve symptoms including nausea, taste disturbance, diarrhea, and epigastric pain. Eradication rates did not significantly differ between the groups (91% vs 79%; p=0.42).

A meta-analysis recently published illustrated that supplementation with *S. boulardii* significantly increased the eradication rate and reduced the risk of overall *H. pylori* therapy-related adverse effects especially diarrhea [57]. Unfortunately, the products used in these studies are not typically sold in the US, which makes selecting a probiotic supported by evidence difficult. Even though a specific strain of *Lactobacillus*

supported by evidence may not be available in the US, it may not be reasonable to extrapolate the effects of that strain to other types of *Lactobacillus* when product selections are limited.

### Probiotics and Inflammatory Bowel Diseases (IBD)

Inflammatory bowel disease (IBD) is a chronic, relapsing, inflammatory disorder of the gastrointestinal tract that includes two entities, namely Crohn's disease (CD) and ulcerative *E. colitis* (UC). Up to one-third of patients with IBD are intolerant of [58] and a further 10% are unresponsive to thiopurine [59]. The role of probiotics in the treatment or relapse prevention in patients with inflammatory bowel diseases (IBD) is more complex and still remains controversial. Anti-tumour necrosis factor (anti-TNF) drugs have proven to be effective in patients with Crohn's disease (CD) [60] and ulcerative *E. colitis* (UC) [61], only approximately one-fifth of all initially treated CD and UC patients are in remission at 1 year. There is still a large gap in the therapeutic armamentarium of both conditions.

Studies using a 16S rRNA technique have shown reductions in *bifidobacteria* [62,63] and *lactobacilli* in patients with UC [64]. *Lactobacillus paracasei* significantly decreased the plasma and lymphocyte content of proinflammatory cytokines in patients with UC [65]. VSL#3 induces IL-10 and downregulates IL-12p40 production by lamina propria DC in patients with UC [66]; similar cytokine changes were seen in patients who were treated with corticosteroids [67]. *E. coli Nissle 1917* seems to have efficacy comparable to that of the antiinflammatory mesalamine for maintenance of remission in ulcerative *E. colitis* patients [68]. In a randomized double-blind trial on patients with active ulcerative *E. colitis*, *E. coli Nissle 1917* did not differ in rate and time to remission compared to placebo [69]. In an open-label trial, VSL#3 did induce remission in 53% and response in 24% over 6 weeks of therapy in patients with active ulcerative *E. colitis* when given along with other treatments [70].

In randomized double-blind, placebo-control trials, *Lactobacillus rhamnosus GG* did not show any superiority over placebo in patients

Trade Name	Manufacturer	Comments
Yo-Plus yogurt	Yoplait Inc	Contains <i>B. animalis</i> laeis Bb-12 in addition to <i>S. thermophilus</i> abd <i>L. bulgaricus</i> per serving.
DanActive Cultured milk	Dannon Inc	Contains <i>S. thermophilus</i> and <i>L. bulgaricus</i> in addition to <i>L. casei</i> DN-114 001. Each serving contains 10 billion CFUs.
VSL#3 packets	Sigma-Tau Pharmaceuticals	Contains <i>B. breve</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>L. acidophilus</i> , <i>L. bulgaricus</i> , <i>L. casei</i> , <i>L. plantarum</i> and <i>Streptococcus thermophiles</i> . Each packet contains 450 billion CFUs. Lemon flavored and is consumed by mixing in atleast 4 oz of cold water.
Philips Colon Health capsules	Proctor & Gamble	Includes <i>Lactobacillus gasseri</i> KS-13, <i>Bifidobacterium bifidum</i> G9-1 and <i>Bifidobacterium longum</i> MM-2. Each capsule contains 1.5 billions cells.
Florastor capsules	Biocodax, inc	Contains <i>Saccharomyces boulardii</i> . Each 250 mg capsule contains 5 billion CFUs.
Florastor Kids	Biocodex Inc	Contains <i>Saccharomyces boulardii</i> . Available as powder.
Attune nutrition bars	Attune Foods	Contains Kosher <i>Lactobacillus acidophilus</i> NCFM, <i>L. casei</i> Lc-11 and <i>Bifidobacterium lactis</i> HN019. Contains 3g fiber. Each serving contains 6.1 billion CFUs.
Align capsules	Proctor & Gamble	Contains <i>Bifidobacterium infantis</i> 35624 in a vegetarian capsule shell. Each capsule contains 1 billion bacteria.
Sustenex	Schiff Nutrition International	Contains Bacillus coagulans GBI-30, 6086 (BC30). Available as capsules, chewies and gummies.
Lactinex	Becton, Dickinson, and Co	Contains <i>Lactobacillus acidophilus</i> and <i>Lactobacillus helveticus (bulgaricus)</i> . Available as capsules and packets.
Fem-Dophilus	Jarrow formulas	Contains <i>Lactobacillus reuteri</i> RC-14, <i>Lactobacillus rhamnosus</i> GR-1. Available as capsules.
Culturelle Digestive	Amerifit Nutrition, Inc	Contains <i>L. rhamnosus</i> GG. Each capsule contains 10 billion CFUs.
Adult Formula CP-1	Custom Probiotics Inc	Contains five probiotic strains: <i>L. Acidophilus</i> , <i>L. Rhamnosus</i> , <i>L. Plantarum</i> , <i>B. Lactis</i> and <i>B. Bifidum</i> . Each capsule has 50 billion CFUs.
OWP probiotics	One Wellness Place	Contains <i>B. longum</i> , <i>B. breves</i> , <i>B. infantis</i> , <i>L. plantarum</i> , <i>L. rhamnosus</i> , and <i>L. acidophilus</i> . Each capsule has 15 billion CFUs.
Good Belly fruit drink	NextFoods	Contains <i>L. plantarum</i> 299v. Each serving contains 20 billion CFUs.

Table 3: List of selected commercially available probiotics in United States.

with active Crohn's disease [71] and patients with Crohn's disease who are in remission with medical therapy [72]. In randomized double-blind, placebo-control trial, *Lactobacillus johnsonii* LA1 did show statistically non-significant decrease in endoscopic recurrence at 6 months when compared with placebo (49% vs 64%) [73].

Routine use of probiotics for IBD may be premature at this stage as we need stronger evidence in the form of large randomized double blinded and placebo control studies and meta-analysis of single probiotic strains to support it.

### Probiotics and Irritable Bowel Syndrome (IBS)

Epidemiological, physiological and clinical studies have suggested the role of intestinal bacteria in the pathogenesis of IBS. Many previous studies indicate that gastroenteritis is a trigger for IBS. In Canada following an outbreak of gastroenteritis, a cohort analysis revealed an increased odd of IBS within 2 years (OR 4.8) [74] and continued for 8 years [75]. In another study, incidence of gastroenteritis in the previous 2 years was associated with almost a four-fold increase in the risk of developing IBS [76]. Physiological studies on animals and humans showed a profound effect of alterations in the composition of the intestinal microbiota (dysbiosis) on the intestinal physiological functions and IBS [77]. A review of several case-control studies revealed abnormal breath test results in patients with IBS following a sugar challenge when compared with controls [78]. The increased risk of developing IBS following gastroenteritis, dysbiosis, and elevated luminal gas production and immune activation, indicate that the gastrointestinal microbiota may be a therapeutic target in IBS.

Though numerous RCTs have evaluated the efficacy of probiotics in IBS patients, most suffer from serious methodological flaws [79]. In a recent systematic review, Brenner and colleagues reported that of 16 RCTs evaluating probiotics in the treatment of IBS, *Bifidobacterium infantis* 35624 was the only probiotic which provided significant improvements in IBS symptoms [80]. In randomized cross-over trials in 59 children with IBS, VSL#3 demonstrated a greater improvement in global symptoms, abdominal pain and abdominal bloating in the probiotic group [81]. Some meta-analysis indicated a more beneficial impact of probiotics on global symptoms than on abdominal pain and flatulence [82-84].

*Bifidobacterium infantis* alone is available in the market as Align capsules or in combination with other probiotic organisms as OWP probiotic capsules, and VSL#3 packets. More evidence is needed before using probiotics for symptom control in IBS.

### Probiotics and Acute Pancreatitis

Probiotics have been shown to be effective in preventing complications in experimental acute pancreatitis by reducing bacterial translocation [85-87]. A clinical trial conducted by Oláh et al. [88] on patients with acute pancreatitis with *L. plantarum* 299 dose of  $1 \times 10^9$  along with oat fiber significantly reduced infected pancreatic necrosis and the number of surgical interventions. Subsequently, several studies reported similarly positive effects of probiotics with or without prebiotics [89-93]. Besselink et al. [94] (PROPATRIA trial) conducted a large multi-center, randomized double-blinded controlled trial involving 296 patients in 15 hospitals, and compared the use of a multi-species probiotics preparation with a placebo. This study showed that infectious complications occurred in 30% of the patients in the probiotics group and in 28% of the placebo group. Nine patients developed bowel ischemia (8 died) in the probiotics group, whereas none developed this complication in the placebo group. Multiple

organ failure occurred in 22% of the patients in the probiotics group and in 10% in the placebo group. In all, 16% patients in the probiotics group and 6% in the placebo group died. Further analyses suggested that higher rates of bowel ischemia in the probiotic patients (6% vs. 0%) may have accounted for the between-group disparity in mortality. This study has been criticized for its design, approval and conduct [95]. The patients in the Besselink group received a higher number and more strains of probiotic organisms (six strains of probiotics vs. 1-4 strains of probiotics in other studies) and some of the patients were receiving pressors. Randomized controlled trials and meta-analysis have not demonstrated significant benefits of prophylactic antibiotics on patients with necrotizing acute pancreatitis [96-98]. Further large-scale, high-quality, placebo-controlled, double-blind trials are needed.

### Probiotics and Necrotizing Enterocolitis (NEC)

Necrotizing enterocolitis is a potentially devastating disease, characterized by severe intestinal inflammation and necrosis, which occurs primarily in preterm infants. The risk of developing NEC is inversely related to gestational age and birth weight. Neonates younger than 28 weeks gestation and of extremely low birth weight (<1000 g) are particularly susceptible. An exaggerated inflammatory response of the immature intestine occurs during a complex interplay between bacterial colonization, initiation of enteral nutrition, and hypoxic-related intestinal injury [99].

Evidence suggests that bacterial colonization patterns are important in the pathogenesis of NEC. Studies have shown that preterm infants of mothers receiving broad-spectrum antibiotics prenatally or preterm infants receiving antibiotics directly postnatally are at higher risk for NEC [100,101]. Isolated studies have demonstrated associations with organisms including *Enterobacteriaceae* [102,103], delta toxin positive methicillin resistant *Staphylococcus aureus* [104], and *Clostridium spp* [105,106]. There are 4 meta-analyses on this subject [107-110]. Two more RCTs have been published since the meta-analyses were completed [111,112]. Each meta-analysis, as well as the 2 recent RCTs, documented reduced rates of NEC and mortality with the use of prophylactic probiotics with an overall reduction in the relative risk (RR) of NEC (Bell  $\geq 2$ ) to 0.35 (95 % CI 0.23-0.55) and of mortality to 0.41 (0.28-0.60) [113]. Best results appear to be achieved with probiotics based on 2 or more probiotic species and/or with a combination of *Bifidobacterium spp.* and *Lactobacillus acidophilus*.

There was an increased risk of sepsis in neonates receiving orally administered probiotics in the randomized, controlled clinical trial by Lin et al. [111], especially in the most vulnerable neonates with birth weights <750 g. However, none of the positive blood cultures grew *Lactobacillus* or *Bifidobacterium spp.*

Our review of studies found that the use of probiotics reduces the occurrence of NEC and death in premature infants born less than 1500 grams. There is insufficient data with regard to the benefits and potential adverse effects in the most at risk infants less than 750 grams at birth.

### Probiotics and Multi-Organ Dysfunction Syndrome (MODS)

Impaired intestinal barrier function has been assumed to play a role in the development of sepsis and multiple organ failure (MOF) in patients with decreased gut perfusion following major surgery, trauma or shock [114,115]. Feeding probiotics (VSL#3) to experimental animals resulted in a normalization of colonic physiologic function

and barrier integrity in conjunction with a reduction in mucosal levels of proinflammatory cytokines [116]. In a prospective study of 25 patients who developed severe SIRS following ICU admission had significantly lower anaerobes in their gut flora and higher counts of pathogenic *Pseudomonas aeruginosa* and *Staphylococcus* spp. in the gut than healthy volunteers [117]. A double-blind, placebo-controlled, randomized design by Alberda et al. [118] to determine the effects of viable probiotics and probiotic sonicates on the development of MODS in critically ill, enterally fed patients showed a significantly larger increase in systemic IgA and IgG concentrations in patients who received viable probiotics than in the patients who received placebo or sonicates ( $P < 0.05$ ). MODS scores were not significantly affected by probiotic treatment. Most of the patients in this study showed a reduction in CRP concentrations over the treatment period, those patients who received viable probiotics had a lesser decline in CRP concentrations than did those patients who received either placebo or bacterial sonicates. Spindler-Vesel et al. [119] also demonstrated reduced infection rates in trauma patients treated with a combination of probiotics and prebiotics.

There are insufficient data to make a recommendation on the use of prebiotics/probiotics/synbiotics in critically ill patients.

### Probiotics and Allergy and Immune Response

Recent research in mucosal immunology demonstrated interactions between microbes and host at an early age even when mucosal barrier and immune system are still immature [120]. Probiotics have been found to enhance the innate immunity and modulate pathogen induced inflammation via toll-like receptor-regulated signaling pathways [121]. The mode of delivery has a great impact on the acquisition of the intestinal bacteria, also beyond the immediate neonatal period. Vaginally born infants and infants born by cesarean section show major differences in cultural microbiota up to 6 months of age [122]. Infants harboring *Bacteroides fragilis* and *Bifidobacterium* species had more circulating immunoglobulin (Ig) A-secreting and IgM-secreting cells. Bacteria in breast milk and microbes potentially present in the amniotic fluid may affect the composition of gut microbiota [123]. Gut microbiota stimulates the TH1, TH3, and T regulatory cells, which can balance the IL-4, IL-5, and IL-13 secreted by TH2 cells in atopic diseases like allergic rhinoconjunctivitis, asthma and atopic eczema [124].

In a randomized double blind placebo-controlled studies of probiotic use, *Lactobacillus GG* or placebo when given to pregnant mothers with a strong family history of eczema, allergic rhinitis or asthma, and to their infants for the first six months after delivery. The frequency of developing atopic dermatitis in the offspring's of pregnant mothers who received *Lactobacillus GG* was significantly reduced by 2, 4, and 7 years, by 50%, 44%, and 36% respectively [125-127]. *Lactobacillus acidophilus* strain was not able to produce same result in a different study suggesting the strain specificity [128]. *Lactobacillus GG* in combination with *B. lactis* during pregnancy and breastfeeding reduced the risk of atopic eczema and allergic sensitization in child [129], whereas a mixture of probiotics (*Lactobacillus GG*, *L. rhamnosus* LC705, *B. breve*, and *Propionibacterium freudenreichii* ssp. *Shermanii* JS) failed to reduce the risk atopic eczema [130], indicates the strain differences and interactions.

*Lactobacillus GG* also shown to increase protective hemagglutinin inhibition titers against the virus with no side effects when treated for 28 days after administration of live-attenuated influenza vaccine in a randomized, double-blind, placebo-controlled pilot study. This

suggests the role of probiotics as a potential adjuvant to improve influenza vaccine [131]. A pilot study involving healthy adults showed higher levels of antityphoid antibodies when *Lactobacillus GG* was given for 10 days before vaccination than those who received placebo [132].

We suggest not using prebiotics, probiotics, or synbiotics for the prevention of any allergic conditions as bibliographical data do not enable any clear conclusion regarding its beneficial effects on the prevention or treatment of allergy. Initial meta-analyses suggest a benefit of probiotics in reducing the development of eczema, but not any other allergic outcome.

### Probiotics and Ventilator Associated Pneumonia (VAP)

Ventilator-associated pneumonia (VAP), defined as pneumonia occurring more than 48 hours after endotracheal intubation, is a leading hospital-acquired infection in the US [133]. Importantly, VAP prolongs the duration of mechanical ventilation, length of stay in the intensive care unit (ICU) and possible recovery of the lung function [134]. Furthermore, patients with VAP may have a 2 to 10 fold higher risk of death compared to mechanically ventilated patients without pneumonia, with crude mortality rates ranging from 24% to 76% [135,136]. The pathogenesis of VAP is complex but typically involves colonization of the aerodigestive tract with pathogenic bacteria, formation of biofilms, and microaspiration of contaminated secretions [137,138]. Rising antibiotic resistance rates have prompted a transition in research efforts from treatment to prevention [135].

In a clinical trial, Forestier et al. [139] using *Lactobacillus casei rhamnosus* strain 35 (Lcr35) demonstrated a decrease in VAP incidence in probiotic group compared to placebo group (2.9% vs 7.5%) along with decreased new gastric colonization by *Pseudomonas aeruginosa* (3% vs 6%) and new *P. aeruginosa* respiratory colonization (5% vs 12%). Knight et al. [140] performed a prospective, randomized, double-blind, placebo controlled trial by randomized patients to receive either Synbiotic 2000 FORTE (n=130) or placebo (n=129) twice a day. The primary endpoint, VAP incidence, was similar between groups (9% of patients in the synbiotic group and 13% of patients in the placebo group;  $p=0.42$ ). Secondary endpoints, including ventilator days, VAP rates per 1000 ventilator days, ICU length of stay, ICU mortality, and hospital mortality did not differ significantly between groups. The only meta-analysis of randomized, controlled clinical trials using probiotics to prevent ventilator-associated pneumonia (VAP) found significant reductions in the incidence of VAP and length of ICU stay [141]. Five studies were included for analysis, including 2 studies described above [139,140]. Mortality, however, was not affected. Morrow et al. [142] recently reported the results of a single-center, double-blind, clinical trial including 146 mechanically ventilated patients, randomized to receive standard care or enteral probiotics (*Lactobacillus rhamnosus*) twice a day. VAP incidence was decreased from 40 to 19% in patients treated with prophylactic probiotic therapy and standard care, respectively ( $P=0.007$ ) which was associated with an estimated number needed to treat to prevent 1 case of VAP as approximately 5 patients. Additionally, a decrease in infections due to *Clostridium difficile* (18.6 vs. 5.8%;  $P=0.02$ ) was reported. Other measures, such as VAP antibiotic days, duration of mechanical ventilation, ICU stay, hospital stay, and total charges, were similar between groups.

In conclusion, we don't have enough data to firmly support the use of probiotic bacteria in the setting of intensive care units. Well-designed multi-center clinical studies with defined mixtures of probiotics and defined endpoints are warranted in this field.

## Probiotics as Commercial Products

Probiotic research and industry have continued to grow from these early observations, and the global market for probiotic ingredients, supplements, and foods amounted to \$21.6 billion in 2010 and are expected to reach \$31.1 billion by 2015 [143]. Probiotics are available in the market under different trade names and varies in different doses and different combinations. Following are the list of some of the commercial probiotics available in the market (Table 3). The list is to give the readers a sense of what is commercially available, not provide recommendations for probiotic strain use. Administration of bacteria-derived probiotics should be separated from antibiotics by at least 2 hours.

Not all probiotics are created equal and the benefits are strain and dose specific. Some formulations also have prebiotics. The choice of probiotic depends upon the health benefit for which it is required. The number of bacteria per serving (CFUs) matter since the administered probiotic is going to be a tiny and transient part of trillions of bacteria already in your gut. The beneficial effects of probiotics cease in 2-4 weeks after stopping administration. This is because the probiotic bacteria stay in our gut only transiently and do not establish permanent residence.

All yogurts sold in the United States are made with the yogurt starter bacteria (*S. thermophilus* and *L. bulgaricus*). Yogurts frequently do not include the levels of bacteria present in the final product on their labels, so the only way to know if a yogurt carries enough of the right type of probiotics to be beneficial is to contact the manufacturer.

We recommend consumers to check with their health care provider before taking probiotics.

## Safety

In the US, probiotics are classified as dietary supplements by the Food and Drug Administration (FDA), thus having less stringent requirements in their demonstration of safety, efficacy, and purity. Specific strains of probiotics fall into the FDA status of generally regarded as safe, while others do not [144]. Generally regarded as safe status only evaluates safety; clinical efficacy is not assessed during this process.

Probiotics are viable organisms with the potential to induce systemic infection in the host. A review of literature by McFarland found 12 cases of *Lactobacillus* probiotic, mostly in children (9 cases) [145]. There are 24 cases of fungemia in patients associated with the probiotic *S. boulardii* [146]. The major risk factors identified were prematurity in infants, chronic disease, immunodeficiency, and/or debilitation. Munoz et al documented 3 patients with *Saccharomyces cerevisiae* fungemia in an ICU associated with *S. boulardii* therapy [147]. Health care providers should change gloves after handling *S. boulardii* powder. Some experts recommend avoiding *S. boulardii* in patients with central venous catheters [148]. Thirty-nine case reports of infection due to *Lactobacillus rhamnosus* GG were reported between 1950 and 2003 by Cannon et al. [149]. In the clinical trials, no reports of bacteremia or fungemia have been associated with probiotic use. All cases of probiotic bacteremia or fungemia have occurred in patients with underlying immune compromise, chronic disease, or debilitation, and no reports have described sepsis related to probiotic use in otherwise healthy persons. Many case reports of probiotic sepsis describe persons with preexisting intestinal pathology, including diarrhea and short intestine. These may be common indications for probiotic use, but would also

be expected to increase the risk of probiotic translocation through the intestinal mucosa.

Secondly, probiotics have the theoretical risk of transfer of antibiotic-resistance genes to pathogenic bacteria. Many *Lactobacillus* strains are naturally resistant to vancomycin, which raises concerns regarding the possible transfer of such resistance to more pathogenic organisms, particularly enterococci and *Staphylococcus aureus*. However, the vancomycin-resistant genes of *Lactobacillus* spp. are chromosomal and, therefore, not readily transferable to other species [150]. The PROPATRIA trial has been discussed in the probiotics and acute pancreatitis section above.

## Challenges

Extrapolation of immunomodulatory effects found in the laboratory and in animal studies with outcomes in human trials presents a difficult challenge. Immunomodulatory effects conferred by *L. plantarum* WCFS1 *in vitro* [151], in animal models [151,152] as well as in humans [153,154] highlight the difficulties of comparing similar effects by a single strain in different contexts. Generally, the discrepancies between *in vitro* and *in vivo* results observed in published trials can be partly explained by the host contribution (genetic factors, different baseline immune functions between individuals, microbiome diversity, differences in the body sites targeted, intra-person variation) as well as environmental factors (diet, stress, etc.) partially controlled by each individual.

Several properties that are thought to be important for the probiotic effect as they can (at least) modify the survival capacity of the strain *in vivo* clearly differ between strains of different or similar species. They include tolerance to acid, bile, and pancreatin; adherence to mucus or to epithelial cells; enzymatic activity; and antibiotic resistance or production of antimicrobial compounds. Several studies have also shown differences in the immunomodulating properties of various probiotics between strains within the same species. For example, Medina et al. [155] showed that different strains of *Bifidobacterium longum* varied greatly in their capacity to induce cytokine production (IL-10, IFN- $\gamma$  and TNF $\alpha$ ) by peripheral blood mononuclear cells and could even drive the immune responses in different directions. Head-to-head comparisons of different strains in human studies are rare. The implications of this strain-specificity are that for commercial products, documentation of health effects must be conducted on the specific strain being sold, one should avoid any extrapolation of positive or negative effects between probiotic strains or products and meta-analysis of the effect of probiotics with different active molecules should also be avoided.

## Conclusion

Probiotics seem to have promising role in shortening duration of infections or decreasing susceptibility to the pathogens. Use of the different strains, dosage, duration of treatment and smaller size of the trials makes interpretation of the available data more difficult. Current evidence also indicates that probiotic effects are strain-specific, they do not act through the same mechanisms nor are all probiotics indicated for the same health conditions. It is currently unknown whether there are optimal probiotic species, doses, and/or formulations. Although the data with probiotics are still far too weak to convince clinicians, the concept is fascinating, and further studies would be more than welcome.

## References

- Antoine JM (2010) Probiotics: beneficial factors of the defence system. Proc Nutr Soc 69: 429-433.
- Kollath W (1953) Nutrition and the tooth system; general review with special reference to vitamins. Dtsch Zahnarzt Z 8: 7-16.
- Lilly DM, Stillwell RH (1965) Probiotics: Growth-promoting Factors Produced by Microorganisms. Science 147: 747-748.
- (2001) Report of a Joint FAO/WHO Expert Consultation on Evaluation of Health and Nutritional Properties of Probiotics in Food Including Powder Milk with Live Lactic Acid Bacteria. "Health and Nutritional Properties of Probiotics in Food including Powder Milk with Live Lactic Acid Bacteria". Food and Agriculture Organization of the United Nations, World Health Organization.
- de Vrese M, Schrezenmeir J (2008) Probiotics, prebiotics, and synbiotics. Adv Biochem Eng Biotechnol 111: 1-66.
- Ohland CL, Macnaughton WK (2010) Probiotic bacteria and intestinal epithelial barrier function. Am J Physiol Gastrointest Liver Physiol 298: G807-819.
- Yan F, Polk DB (2006) Probiotics as functional food in the treatment of diarrhea. Curr Opin Clin Nutr Metab Care 9: 717-721.
- Mack DR, Ahrne S, Hyde L, Wei S, Hollingsworth MA (2003) Extracellular MUC3 mucin secretion follows adherence of *Lactobacillus* strains to intestinal epithelial cells *in vitro*. Gut 52: 827-833.
- Mattar AF, Teitelbaum DH, Drongowski RA, Yongyi F, Harmon CM, et al. (2002) Probiotics up-regulate MUC-2 mucin gene expression in a Caco-2 cell-culture model. Pediatr Surg Int 18: 586-590.
- Gaudier E, Michel C, Segain JP, Cherbut C, Hoebler C (2005) The VSL# 3 probiotic mixture modifies microflora but does not heal chronic dextran-sodium sulfate-induced colitis or reinforce the mucus barrier in mice. J Nutr 135: 2753-2761.
- Caballero-Franco C, Keller K, De Simone C, Chadee K (2007) The VSL#3 probiotic formula induces mucin gene expression and secretion in colonic epithelial cells. Am J Physiol Gastrointest Liver Physiol 292: G315-322.
- Kelsall BL (2008) Innate and adaptive mechanisms to control [corrected] pathological intestinal inflammation. J Pathol 214: 242-259.
- Möndel M, Schroeder BO, Zimmermann K, Huber H, Nuding S, et al. (2009) Probiotic *E. coli* treatment mediates antimicrobial human beta-defensin synthesis and fecal excretion in humans. Mucosal Immunol 2: 166-172.
- Schlee M, Harder J, Köten B, Stange EF, Wehkamp J, et al. (2008) Probiotic lactobacilli and VSL#3 induce enterocyte beta-defensin 2. Clin Exp Immunol 151: 528-535.
- Penner R, Fedorak RN, Madsen KL (2005) Probiotics and nutraceuticals: non-medical treatments of gastrointestinal diseases. Curr Opin Pharmacol 5: 596-603.
- Alakomi HL, Skyttä E, Saarela M, Mattila-Sandholm T, Latva-Kala K, et al. (2000) Lactic acid permeabilizes gram-negative bacteria by disrupting the outer membrane. Appl Environ Microbiol 66: 2001-2005.
- Liévin-Le Moal V, Servin AL (2006) The front line of enteric host defense against unwelcome intrusion of harmful microorganisms: mucins, antimicrobial peptides, and microbiota. Clin Microbiol Rev 19: 315-337.
- Duquesne S, Petit V, Peduzzi J, Rebuffat S (2007) Structural and functional diversity of microcins, gene-encoded antibacterial peptides from enterobacteria. J Mol Microbiol Biotechnol 13: 200-209.
- Johnson-Henry KC, Hagen KE, Gordonpour M, Tompkins TA, Sherman PM (2007) Surface-layer protein extracts from *Lactobacillus helveticus* inhibit enterohaemorrhagic *Escherichia coli* O157:H7 adhesion to epithelial cells. Cell Microbiol 9: 356-367.
- Wu X, Vallance BA, Boyer L, Bergstrom KS, Walker J, et al. (2008) *Saccharomyces boulardii* ameliorates *Citrobacter rodentium*-induced colitis through actions on bacterial virulence factors. Am J Physiol Gastrointest Liver Physiol 294: G295-306.
- Galdeano CM, Perdígón G (2006) The probiotic bacterium *Lactobacillus casei* induces activation of the gut mucosal immune system through innate immunity. Clin Vaccine Immunol 13: 219-226.
- Ogawa M, Shimizu K, Nomoto K, Takahashi M, Watanuki M, et al. (2001) Protective effect of *Lactobacillus casei* strain Shirota on Shiga toxin-producing *Escherichia coli* O157:H7 infection in infant rabbits. Infect Immun 69: 1101-1108.
- Tien MT, Girardin SE, Regnault B, Le Bourhis L, Dillies MA, et al. (2006) Anti-inflammatory effect of *Lactobacillus casei* on Shigella-infected human intestinal epithelial cells. J Immunol 176: 1228-1237.
- Miller MB, Bassler BL (2001) Quorum sensing in bacteria. Annu Rev Microbiol 55: 165-199.
- Kendall MM, Sperandio V (2007) Quorum sensing by enteric pathogens. Curr Opin Gastroenterol 23: 10-15.
- Medellin-Peña MJ, Wang H, Johnson R, Anand S, Griffiths MW (2007) Probiotics affect virulence-related gene expression in *Escherichia coli* O157:H7. Appl Environ Microbiol 73: 4259-4267.
- Bartlett JG (1996) Management of *Clostridium difficile* infection and other antibiotic-associated diarrhoeas. Eur J Gastroenterol Hepatol 8: 1054-1061.
- Mueller S, Saunier K, Hanisch C, Norin E, Alm L, et al. (2006) Differences in fecal microbiota in different European study populations in relation to age, gender, and country: a cross-sectional study. Appl Environ Microbiol 72: 1027-1033.
- Guigoz Y, Doré J, Schiffrin EJ (2008) The inflammatory status of old age can be nurtured from the intestinal environment. Curr Opin Clin Nutr Metab Care 11: 13-20.
- Szajewska H, Rusczyński M, Radzikowski A (2006) Probiotics in the prevention of antibiotic-associated diarrhea in children: a meta-analysis of randomized controlled trials. J Pediatr 149: 367-372.
- Adam J, Barret C, Barret-Bellet A (1977) Controlled double-blind clinical trials of Ultra-Levure: Multicentre study by 25 physicians in 388 cases. Gazette Medicale de France 84: 2072-2078.
- Surawicz CM, Elmer GW, Speelman P, McFarland LV, Chinn J, et al. (1989) Prevention of antibiotic-associated diarrhea by *Saccharomyces boulardii*: a prospective study. Gastroenterology 96: 981-988.
- McFarland LV, Surawicz CM, Greenberg RN, Fekety R, Elmer GW, et al. (1994) A randomized placebo-controlled trial of *Saccharomyces boulardii* in combination with standard antibiotics for *Clostridium difficile* disease. JAMA 271: 1913-1918.
- Kotowska M, Albrecht P, Szajewska H (2005) *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhoea in children: a randomized double-blind placebo-controlled trial. Aliment Pharmacol Ther 21: 583-590.
- Lewis SJ, Potts LF, Barry RE (1998) The lack of therapeutic effect of *Saccharomyces boulardii* in the prevention of antibiotic-related diarrhoea in elderly patients. J Infect 36: 171-174.
- Can M, Besirbellioglu BA, Avci IY, Beker CM, Pahsa A (2006) Prophylactic *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhea: a prospective study. Med Sci Monit 12: P119-22.
- McFarland LV (2010) Systematic review and meta-analysis of *Saccharomyces boulardii* in adult patients. World J Gastroenterol 16: 2202-2222.
- Szajewska H, Mrukowicz J (2005) Meta-analysis: non-pathogenic yeast *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhoea. Aliment Pharmacol Ther 22: 365-372.
- Wenus C, Goll R, Loken EB, Biong AS, Halvorsen DS, et al. (2008) Prevention of antibiotic-associated diarrhoea by a fermented probiotic milk drink. Eur J Clin Nutr 62: 299-301.
- Vanderhoof JA, Whitney DB, Antonson DL, Hanner TL, Lupo JV, et al. (1999) *Lactobacillus GG* in the prevention of antibiotic-associated diarrhea in children. J Pediatr 135: 564-568.
- Arvola T, Laiho K, Torkkeli S, Mykkänen H, Salminen S, et al. (1999) Prophylactic *Lactobacillus GG* reduces antibiotic-associated diarrhea in children with respiratory infections: a randomized study. Pediatrics 104: e64.
- Thomas MR, Litin SC, Osmon DR, Corr AP, Weaver AL, et al. (2001) Lack of effect of *Lactobacillus GG* on antibiotic-associated diarrhea: a randomized, placebo-controlled trial. Mayo Clin Proc 76: 883-889.
- Castagliuolo I, LaMont JT, Nikulasson ST, Pothoulakis C (1996) *Saccharomyces boulardii* protease inhibits *Clostridium difficile* toxin A effects in the rat ileum. Infect Immun 64: 5225-5232.

44. Qamar A, Aboudola S, Warny M, Michetti P, Pothoulakis C, et al. (2001) *Saccharomyces boulardii* stimulates intestinal immunoglobulin A immune response to *Clostridium difficile* toxin A in mice. *Infect Immun* 69: 2762-2765.
45. McFarland LV, Surawicz CM, Greenberg RN, Elmer GW, Moyer KA, et al. (1995) Prevention of beta-lactam-associated diarrhea by *Saccharomyces boulardii* compared with placebo. *Am J Gastroenterol* 90: 439-448.
46. Plummer S, Weaver MA, Harris JC, Dee P, Hunter J (2004) *Clostridium difficile* pilot study: effects of probiotic supplementation on the incidence of *C. difficile* diarrhoea. *Int Microbiol* 7: 59-62.
47. Thomas MR, Litin SC, Osmon DR, Corr AP, Weaver AL, et al. (2001) Lack of effect of *Lactobacillus GG* on antibiotic-associated diarrhea: a randomized, placebo-controlled trial. *Mayo Clin Proc* 76: 883-889.
48. Surawicz CM, Elmer GW, Speelman P, McFarland LV, Chinn J, et al. (1989) Prevention of antibiotic-associated diarrhea by *Saccharomyces boulardii*: a prospective study. *Gastroenterology* 96: 981-988.
49. Lewis SJ, Potts LF, Barry RE (1998) The lack of therapeutic effect of *Saccharomyces boulardii* in the prevention of antibiotic-related diarrhoea in elderly patients. *J Infect* 36: 171-174.
50. Kotowska M, Albrecht P, Szajewska H (2005) *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhoea in children: a randomized double-blind placebo-controlled trial. *Aliment Pharmacol Ther* 21: 583-590.
51. Can M, Besirbellioglu BA, Avci IY, Beker CM, Pahsa A (2006) Prophylactic *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhea: a prospective study. *Med Sci Monit* 12: P19-22.
52. Hickson M, D'Souza AL, Muthu N, Rogers TR, Want S, et al. (2007) Use of probiotic *Lactobacillus* preparation to prevent diarrhoea associated with antibiotics: randomised double blind placebo controlled trial. *BMJ* 335: 80.
53. Surawicz CM, McFarland LV, Greenberg RN, Rubin M, Fekety R, et al. (2000) The search for a better treatment for recurrent *Clostridium difficile* disease: use of high-dose vancomycin combined with *Saccharomyces boulardii*. *Clin Infect Dis* 31: 1012-1017.
54. Armuzzi A, Cremonini F, Bartolozzi F, Canducci F, Candelli M, et al. (2001) The effect of oral administration of *Lactobacillus GG* on antibiotic-associated gastrointestinal side-effects during *Helicobacter pylori* eradication therapy. *Aliment Pharmacol Ther* 15: 163-169.
55. Cremonini F, Di Caro S, Covino M, Armuzzi A, Gabrielli M, et al. (2002) Effect of different probiotic preparations on anti-*helicobacter pylori* therapy-related side effects: a parallel group, triple blind, placebo-controlled study. *Am J Gastroenterol* 97: 2744-2749.
56. Myllyluoma E, Veijola L, Ahlroos T, Tynkkynen S, Kankuri E, et al. (2005) Probiotic supplementation improves tolerance to *Helicobacter pylori* eradication therapy—a placebo-controlled, double-blind randomized pilot study. *Aliment Pharmacol Ther* 21: 1263-1272.
57. Szajewska H, Horvath A, Piwowarczyk A (2010) Meta-analysis: the effects of *Saccharomyces boulardii* supplementation on *Helicobacter pylori* eradication rates and side effects during treatment. *Aliment Pharmacol Ther* 32: 1069-1079.
58. Fraser AG, Orchard TR, Jewell DP (2002) The efficacy of azathioprine for the treatment of inflammatory bowel disease: a 30 year review. *Gut* 50: 485-489.
59. Candy S, Wright J, Gerber M, Adams G, Gerig M, et al. (1995) A controlled double blind study of azathioprine in the management of Crohn's disease. *Gut* 37: 674-678.
60. Hanauer SB, Feagan BG, Lichtenstein GR, Mayer LF, Schreiber S, et al. (2002) Maintenance infliximab for Crohn's disease: the ACCENT I randomised trial. *Lancet* 359: 1541-1549.
61. Rutgeerts P, Sandborn WJ, Feagan BG, Reinisch W, Olson A, et al. (2005) Infliximab for induction and maintenance therapy for ulcerative colitis. *N Engl J Med* 353: 2462-2476.
62. Mylonaki M, Rayment NB, Rampton DS, Hudspith BN, Brostoff J (2005) Molecular characterization of rectal mucosa-associated bacterial flora in inflammatory bowel disease. *Inflamm Bowel Dis* 11: 481-487.
63. Macfarlane S, Furrie E, Kennedy A, Cummings JH, Macfarlane GT (2005) Mucosal bacteria in ulcerative colitis. *Br J Nutr* 93 Suppl 1: S67-72.
64. Bullock NR, Booth JC, Gibson GR (2004) Comparative composition of bacteria in the human intestinal microflora during remission and active ulcerative colitis. *Curr Issues Intest Microbiol* 5: 59-64.
65. Federico A, Tuccillo C, Grossi E, Abbiati R, Garbagna N, et al. (2009) The effect of a new symbiotic formulation on plasma levels and peripheral blood mononuclear cell expression of some pro-inflammatory cytokines in patients with ulcerative colitis: a pilot study. *Eur Rev Med Pharmacol Sci* 13: 285-293.
66. Ng SC, Plamondon S, Al-Hassi HO, Kamm MA, Knight SC, et al. (2008) M1202 Effective probiotic treatment (VSL#3), but not placebo, in acute ulcerative colitis is associated with downregulation of inflammatory intestinal dendritic cells. *Gastroenterology* 134.
67. Ng SC, Plamondon S, Al-Hassi HO, Kamm MA, Knight SC, et al. (2008) Corticosteroids increase interleukin-10 and inhibit interleukin-12p40 production by intestinal dendritic cells in acute ulcerative colitis: novel mechanisms of therapy. *Gastroenterology* 134.
68. Kruis W, Fric P, Pokrotnieks J, Lukás M, Fixa B, et al. (2004) Maintaining remission of ulcerative colitis with the probiotic *Escherichia coli* Nissle 1917 is as effective as with standard mesalazine. *Gut* 53: 1617-1623.
69. Rembacken BJ, Snelling AM, Hawkey PM, Chalmers DM, Axon AT (1999) Non-pathogenic *Escherichia coli* versus mesalazine for the treatment of ulcerative colitis: a randomised trial. *Lancet* 354: 635-639.
70. Bibiloni R, Fedorak RN, Tannock GW, Madsen KL, Gionchetti P, et al. (2005) VSL#3 probiotic-mixture induces remission in patients with active ulcerative colitis. *Am J Gastroenterol* 100: 1539-1546.
71. Schultz M, Timmer A, Herfarth HH, Sartor RB, Vanderhoof JA, et al. (2004) *Lactobacillus GG* in inducing and maintaining remission of Crohn's disease. *BMC Gastroenterol* 4: 5.
72. Bousvaros A, Guandalini S, Baldassano RN, Botelho C, Evans J, et al. (2005) A randomized, double-blind trial of *Lactobacillus GG* versus placebo in addition to standard maintenance therapy for children with Crohn's disease. *Inflamm Bowel Dis* 11: 833-839.
73. Marteau P, Lémann M, Seksik P, Laharie D, Colombel JF, et al. (2006) Ineffectiveness of *Lactobacillus johnsonii* LA1 for prophylaxis of postoperative recurrence in Crohn's disease: a randomised, double blind, placebo controlled GETAID trial. *Gut* 55: 842-847.
74. Marshall JK, Thabane M, Garg AX, Clark WF, Salvadori M, et al. (2006) Incidence and epidemiology of irritable bowel syndrome after a large waterborne outbreak of bacterial dysentery. *Gastroenterology* 131: 445-450.
75. Marshall JK, Thabane M, Garg AX, Clark WF, Moayyedi P, et al. (2010) Eight year prognosis of postinfectious irritable bowel syndrome following waterborne bacterial dysentery. *Gut* 59: 605-611.
76. Porter CK, Gormley R, Tribble DR, Cash BD, Riddle MS (2011) The Incidence and gastrointestinal infectious risk of functional gastrointestinal disorders in a healthy US adult population. *Am J Gastroenterol* 106: 130-138.
77. Husebye E, Hellström PM, Sundler F, Chen J, Midtvedt T (2001) Influence of microbial species on small intestinal myoelectric activity and transit in germ-free rats. *Am J Physiol Gastrointest Liver Physiol* 280: G368-380.
78. Shah ED, Basseri RJ, Chong K, Pimentel M (2010) Abnormal breath testing in IBS: a meta-analysis. *Dig Dis Sci* 55: 2441-2449.
79. Moayyedi P, Ford AC, Talley NJ, Cremonini F, Foxx-Orenstein AE, et al. (2010) The efficacy of probiotics in the treatment of irritable bowel syndrome: a systematic review. *Gut* 59: 325-332.
80. Brenner DM, Moeller MJ, Chey WD, Schoenfeld PS (2009) The utility of probiotics in the treatment of irritable bowel syndrome: a systematic review. *Am J Gastroenterol* 104: 1033-1049.
81. Guandalini S, Magazzù G, Chiaro A, La Balestra V, Di Nardo G, et al. (2010) VSL#3 improves symptoms in children with irritable bowel syndrome: a multicenter, randomized, placebo-controlled, double-blind, crossover study. *J Pediatr Gastroenterol Nutr* 51: 24-30.
82. McFarland LV, Dublin S (2008) Meta-analysis of probiotics for the treatment of irritable bowel syndrome. *World J Gastroenterol* 14: 2650-2661.
83. National Institute for Health and Clinical Excellence (NICE) (2008). Irritable bowel syndrome in primary care: diagnosis and management of irritable bowel syndrome in primary care. National Collaborating Centre for Nursing and Supportive Care.
84. Hoveyda N, Heneghan C, Mahtani KR, Perera R, Roberts N, et al. (2009) A

- systematic review and meta-analysis: probiotics in the treatment of irritable bowel syndrome. *BMC Gastroenterol* 9: 15.
85. Akyol S, Mas MR, Comert B, Ateskan U, Yasar M, et al. (2003) The effect of antibiotic and probiotic combination therapy on secondary pancreatic infections and oxidative stress parameters in experimental acute necrotizing pancreatitis. *Pancreas* 26: 363-367.
86. Muftuoglu MA, Isikgor S, Tosun S, Saglam A (2006) Effects of probiotics on the severity of experimental acute pancreatitis. *Eur J Clin Nutr* 60: 464-468.
87. van Minnen LP, Timmerman HM, Lutgendorff F, Verheem A, Harmsen W, et al. (2007) Modification of intestinal flora with multispecies probiotics reduces bacterial translocation and improves clinical course in a rat model of acute pancreatitis. *Surgery* 141: 470-480.
88. Oláh A, Belágyi T, Issekutz A, Gamal ME, Bengmark S (2002) Randomized clinical trial of specific lactobacillus and fibre supplement to early enteral nutrition in patients with acute pancreatitis. *Br J Surg* 89: 1103-1107.
89. Qin HL, Zheng JJ, Tong DN, Chen WX, Fan XB, et al. (2008) Effect of *Lactobacillus plantarum* enteral feeding on the gut permeability and septic complications in the patients with acute pancreatitis. *Eur J Clin Nutr* 62: 923-930.
90. Oláh A, Belágyi T, Pótó L, Romics L Jr, Bengmark S (2007) Synbiotic control of inflammation and infection in severe acute pancreatitis: a prospective, randomized, double blind study. *Hepatogastroenterology* 54: 590-594.
91. Li YM (2007) Adjuvant therapy for probiotics in patients with severe acute pancreatitis: An analysis of 14 cases. *World Chinese Journal of Digestology* 15: 302-304.
92. Karakan T, Ergun M, Dogan I, Cindoruk M, Unal S (2007) Comparison of early enteral nutrition in severe acute pancreatitis with prebiotic fiber supplementation versus standard enteral solution: a prospective randomized double-blind study. *World J Gastroenterol* 13: 2733-2737.
93. Wu XG, Zhang QC (2009) Adjuvant therapy for probiotics in patients with severe acute pancreatitis with hepatic lesion: an analysis of 27 cases. *Clin Med* 29: 51-52.
94. Besselink MG, van Santvoort HC, Buskens E, Boermeester MA, van Goor H, et al. (2008) Probiotic prophylaxis in predicted severe acute pancreatitis: a randomized, double-blind, placebo-controlled trial. *Lancet* 371: 651-659.
95. Sheldon T (2010) Dutch probiotics study is criticised for its "design, approval, and conduct". *BMJ* 340: c77.
96. Dellinger EP, Tellado JM, Soto NE, Ashley SW, Barie PS, et al. (2007) Early antibiotic treatment for severe acute necrotizing pancreatitis: a randomized, double-blind, placebo-controlled study. *Ann Surg* 245: 674-683.
97. Isenmann R, Rünzi M, Kron M, Kahl S, Kraus D, et al. (2004) Prophylactic antibiotic treatment in patients with predicted severe acute pancreatitis: a placebo-controlled, double-blind trial. *Gastroenterology* 126: 997-1004.
98. Mazaki T, Ishii Y, Takayama T (2006) Meta-analysis of prophylactic antibiotic use in acute necrotizing pancreatitis. *Br J Surg* 93: 674-684.
99. Hsueh W, Caplan MS, Qu XW, Tan XD, De Plaen IG, et al. (2003) Neonatal necrotizing enterocolitis: clinical considerations and pathogenetic concepts. *Pediatr Dev Pathol* 6: 6-23.
100. Cotten CM, Taylor S, Stoll B, Goldberg RN, Hansen NI, et al. (2009) Prolonged duration of initial empirical antibiotic treatment is associated with increased rates of necrotizing enterocolitis and death for extremely low birth weight infants. *Pediatrics* 123: 58-66.
101. Kenyon SL, Taylor DJ, Tarnow-Mordi W; ORACLE Collaborative Group (2001) Broad-spectrum antibiotics for preterm, prelabour rupture of fetal membranes: the ORACLE I randomised trial. *Lancet* 357: 979-988.
102. Bell MJ, Shackelford PG, Feigin RD, Ternberg JL, Brotherton T (1979) Alterations in gastrointestinal microflora during antimicrobial therapy for necrotizing enterocolitis. *Pediatrics* 63: 425-428.
103. Millar MR, MacKay P, Levene M, Langdale V, Martin C (1992) *Enterobacteriaceae* and neonatal necrotizing enterocolitis. *Arch Dis Child* 67: 53-56.
104. Overturf GD, Sherman MP, Scheifele DW, Wong LC (1990) Neonatal necrotizing enterocolitis associated with delta toxin-producing methicillin-resistant *Staphylococcus aureus*. *Pediatr Infect Dis J* 9: 88-91.
105. Sturm R, Staneck JL, Sauffer LR, Neblett 3rd WW (1980) Neonatal necrotizing enterocolitis associated with penicillin-resistant, toxigenic *Clostridium butyricum*. *Pediatrics* 66: 928-931.
106. Blakey JL, Lubitz L, Campbell NT, Gillam GL, Bishop RF, et al. (1985) Enteric colonization in sporadic neonatal necrotizing enterocolitis. *J Pediatr Gastroenterol Nutr* 4: 591-595.
107. Alfaleh K, Bassler D (2011) Probiotics for prevention of necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev* CD005496.
108. Deshpande G, Rao S, Patole S (2007) Probiotics for prevention of necrotizing enterocolitis in preterm neonates with very low birthweight: a systematic review of randomised controlled trials. *Lancet* 369: 1614-1620.
109. Mihatsch WA (2008) Probiotika bei Frühgeborenen. *Monatsschr Kinderheilk* 156: 1070-1075.
110. Hammerman C, Kaplan M (2006) Probiotics and neonatal intestinal infection. *Curr Opin Infect Dis* 19: 277-282.
111. Lin HC, Hsu CH, Chen HL, Chung MY, Hsu JF, et al. (2008) Oral probiotics prevent necrotizing enterocolitis in very low birth weight preterm infants: a multicenter, randomized, controlled trial. *Pediatrics* 122: 693-700.
112. Samanta M, Sarkar M, Ghosh P, Ghosh Jk, Sinha Mk, et al. (2009) Prophylactic probiotics for prevention of necrotizing enterocolitis in very low birth weight newborns. *J Trop Pediatr* 55: 128-131.
113. Guthmann F, Kluthe C, Bühner C (2010) Probiotics for prevention of necrotizing enterocolitis: an updated meta-analysis. *Klin Padiatr* 222: 284-290.
114. Derikx JP, Poeze M, van Bijnen AA, Buurman WA, Heineman E (2007) Evidence for intestinal and liver epithelial cell injury in the early phase of sepsis. *Shock* 28: 544-548.
115. Holland J, Carey M, Hughes N, Sweeney K, Byrne PJ, et al. (2005) Intraoperative splanchnic hypoperfusion, increased intestinal permeability, down-regulation of monocyte class II major histocompatibility complex expression, exaggerated acute phase response, and sepsis. *Am J Surg* 190: 393-400.
116. Madsen K, Cornish A, Soper P, McKaigney C, Jijon H, et al. (2001) Probiotic bacteria enhance murine and human intestinal epithelial barrier function. *Gastroenterology* 121: 580-591.
117. Shimizu K, Ogura H, Goto M, Asahara T, Nomoto K, et al. (2006) Altered gut flora and environment in patients with severe SIRS. *J Trauma* 60: 126-133.
118. Alberda C, Gramlich L, Meddings J, Field C, McCargar L, et al. (2007) Effects of probiotic therapy in critically ill patients: a randomized, double-blind, placebo-controlled trial. *Am J Clin Nutr* 85: 816-823.
119. Spindler-Vesela A, Bengmark S, Vovk I, Cerovic O, Kompan L (2007) Synbiotics, prebiotics, glutamine, or peptide in early enteral nutrition: a randomized study in trauma patients. *JPEN J Parenter Enteral Nutr* 31: 119-126.
120. Rook GA, Brunet LR (2005) Microbes, immunoregulation, and the gut. *Gut* 54: 317-320.
121. Vanderpool C, Yan F, Polk DB (2008) Mechanisms of probiotic action: Implications for therapeutic applications in inflammatory bowel diseases. *Inflamm Bowel Dis* 14: 1585-1596.
122. Grönlund MM, Arvilommi H, Kero P, Lehtonen OP, Isolauri E (2000) Importance of intestinal colonisation in the maturation of humoral immunity in early infancy: a prospective follow up study of healthy infants aged 0-6 months. *Arch Dis Child Fetal Neonatal Ed* 83: F186-192.
123. Martín R, Langa S, Reviriego C, Jiménez E, Marín ML, et al. (2003) Human milk is a source of lactic acid bacteria for the infant gut. *J Pediatr* 143: 754-758.
124. Isolauri E, Huurre A, Salminen S, Impivaara O (2004) The allergy epidemic extends beyond the past few decades. *Clin Exp Allergy* 34: 1007-1010.
125. Kalliomäki M, Salminen S, Arvilommi H, Kero P, Koskinen P, et al. (2001) Probiotics in primary prevention of atopic disease: a randomized placebo-controlled trial. *Lancet* 357: 1076-1079.
126. Kalliomäki M, Salminen S, Poussa T, Arvilommi H, Isolauri E (2003) Probiotics and prevention of atopic disease: 4-year follow-up of a randomized placebo-controlled trial. *Lancet* 361: 1869-1871.
127. Kalliomäki M, Salminen S, Poussa T, Isolauri E (2007) Probiotics during the

- first 7 years of life: a cumulative risk reduction of eczema in a randomized, placebo-controlled trial. *J Allergy Clin Immunol* 119: 1019-1021.
128. Taylor AL, Hale J, Wiltschut J, Lehmann H, Dunstan JA, et al. (2006) Effects of probiotic supplementation for the first 6 months of life on allergen- and vaccine-specific immune responses. *Clin Exp Allergy* 36: 1227-1235.
129. Huurre A, Laitinen K, Rautava S, Korkeamäki M, Isolauri E (2008) Impact of maternal atopy and probiotic supplementation during pregnancy on infant sensitization: a double-blind placebo-controlled study. *Clin Exp Allergy* 38: 1342-1348.
130. Viljanen M, Savilahti E, Haahtela T, Juntunen-Backman K, Korpela R, et al. (2005) Probiotics in the treatment of atopic eczema/dermatitis syndrome in infants: a double-blind placebo-controlled trial. *Allergy* 60: 494-500.
131. Davidson LE, Fiorino AM, Snyderman DR, Hibberd PL (2011) *Lactobacillus GG* as an immune adjuvant for live-attenuated influenza vaccine in healthy adults: a randomized double-blind placebo-controlled trial. *Eur J Clin Nutr* 65: 501-507.
132. Teitelbaum JE, Walker WA (2002) Nutritional impact of pre- and probiotics as protective gastrointestinal organisms. *Annu Rev Nutr* 22: 107-138.
133. American Thoracic Society, Infectious Diseases Society of America (2005) Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *Am J Respir Crit Care Med* 171: 388-416.
134. Safdar N, Dezfulian C, Collard HR, Saint S (2005) Clinical and economic consequences of ventilator-associated pneumonia: a systematic review. *Crit Care Med* 33: 2184-2193.
135. Craven DE, De Rosa FG, Thornton D (2002) Nosocomial pneumonia: emerging concepts in diagnosis, management, and prophylaxis. *Curr Opin Crit Care* 8: 421-429.
136. Chastre J, Fagon JY (2002) Ventilator-associated pneumonia. *Am J Respir Crit Care Med* 165: 867-903.
137. Kollef MH (2005) What is ventilator-associated pneumonia and why is it important? *Respir Care* 50: 714-721.
138. Kollef MH (2004) Prevention of hospital-associated pneumonia and ventilator-associated pneumonia. *Crit Care Med* 32: 1396-1405.
139. Forestier C, Guelon D, Cluytens V, Gillart T, Sirot J, et al. (2008) Oral probiotic and prevention of *Pseudomonas aeruginosa* infections: a randomized, double-blind, placebo-controlled pilot study in intensive care unit patients. *Crit Care* 12: R69.
140. Knight DJ, Gardiner D, Banks A, Snape SE, Weston VC, et al. (2009) Effect of synbiotic therapy on the incidence of ventilator associated pneumonia in critically ill patients: a randomised, double-blind, placebo-controlled trial. *Intensive Care Med* 35: 854-861.
141. Siempos II, Ntaidou TK, Falagas ME (2010) Impact of the administration of probiotics on the incidence of ventilator-associated pneumonia: a meta-analysis of randomized controlled trials. *Crit Care Med* 38: 954-962.
142. Morrow LE, Kollef MH, Casale TB (2010) Probiotic prophylaxis of ventilator-associated pneumonia: a blinded, randomized, controlled trial. *Am J Respir Crit Care Med* 182: 1058-1064.
143. <http://www.bccresearch.com/report/probiotic-supplements-food-fod035b.html>
144. Liong MT (2008) Safety of probiotics: translocation and infection. *Nutr Rev* 66: 192-202.
145. McFarland LV (2009) Evidence-based review of probiotics for antibiotic-associated diarrhea and *Clostridium difficile* infections. *Anaerobe* 15: 274-280.
146. Boyle RJ, Robins-Browne RM, Tang ML (2006) Probiotic use in clinical practice: what are the risks? *Am J Clin Nutr* 83: 1256-1264.
147. Muñoz P, Bouza E, Cuenca-Estrella M, Eiros JM, Pérez MJ, et al. (2005) *Saccharomyces cerevisiae* fungemia: an emerging infectious disease. *Clin Infect Dis* 40: 1625-1634.
148. Buts JP, Bernasconi P (2005) *Saccharomyces boulardii*: basic science and clinical applications in gastroenterology. *Gastroenterol Clin North Am* 34: 515-532, x.
149. Cannon JP, Lee TA, Bolanos JT, Danziger LH (2005) Pathogenic relevance of *Lactobacillus*: a retrospective review of over 200 cases. *Eur J Clin Microbiol Infect Dis* 24: 31-40.
150. Tynkkynen S, Singh KV, Varmanen P (1998) Vancomycin resistance factor of *Lactobacillus rhamnosus GG* in relation to enterococcal vancomycin resistance (van) genes. *Int J Food Microbiol* 41: 195-204.
151. Granette C, Nutten S, Palumbo E, Morath S, Hermann C, et al. (2005) Enhanced antiinflammatory capacity of a *Lactobacillus plantarum* mutant synthesizing modified teichoic acids. *Proc Natl Acad Sci U S A* 102: 10321-10326.
152. Marco ML, Bongers RS, de Vos WM, Kleerebezem M (2007) Spatial and temporal expression of *Lactobacillus plantarum* genes in the gastrointestinal tracts of mice. *Appl Environ Microbiol* 73: 124-132.
153. van Baarlen P, Troost FJ, van Hemert S, van der Meer C, de Vos WM, et al. (2009) Differential NF-kappaB pathways induction by *Lactobacillus plantarum* in the duodenum of healthy humans correlating with immune tolerance. *Proc Natl Acad Sci U S A* 106: 2371-2376.
154. Troost FJ, van Baarlen P, Lindsey P, Kodde A, de Vos WM, et al. (2008) Identification of the transcriptional response of human intestinal mucosa to *Lactobacillus plantarum* WCFS1 *in vivo*. *BMC Genomics* 9: 374.
155. Medina M, Izquierdo E, Ennahar S, Sanz Y (2007) Differential immunomodulatory properties of *Bifidobacterium* logum strains: relevance to probiotic selection and clinical applications. *Clin Exp Immunol* 150: 531-538.

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